

# Tumor Anti-angiogenesis: Nano-Arsenal will Kill the Crab

Feroz Alam<sup>1,✉</sup>, Kafil Akhtar<sup>1</sup>, Veena Maheshwari<sup>1</sup>, Mahboob Hasan<sup>1</sup>

<sup>1</sup>Department of Pathology, Jawaharlal Nehru Medical College, Aligarh Muslim University (AMU), Aligarh, Uttar Pradesh, India

## Abstract

Angiogenesis: the process of developing new blood vessels from the pre-existing ones is a very crucial process in tumor formation, growth, progression and metastasis. The central role of angiogenesis in tumor growth and progression compels targeting tumor vasculature as a therapeutic mean. Nanoparticles have revolutionized the domain of cancer therapeutics, molecularly targeted anti-cancer drugs represents one of the most significant recent advances in clinical oncology. Various types of nano-drug complexes which targets tumor vasculature preferentially, are being increasingly tested for optimal therapeutic efficacy and minimal side-effects. Here, we review some of these anti-angiogenic nano-drug complexes.

## Keywords

Anti-angiogenesis, cancer treatment, nanotechnology, active targeting

## Introduction

During embryogenesis, endothelial progenitor cells (angioblasts) form a primitive vascular network of small capillaries, a process termed “vasculogenesis.” Further transformations of this vascular network proceed during “angiogenesis” when new vessels are formed from already existing ones. During angiogenesis, the primary vascular network significantly expands due to capillary branching and is transformed into the highly organized vascular net<sup>1</sup>. Later, the vascular network matures, and capillaries fuse to form bigger vessels, arteries, and veins. Vasculature in healthy adult is very stable and with the exception of few rare events such as cyclical growth of vessels in the ovarian corpus luteum or during pregnancy, angiogenic activities are rare in adult individuals<sup>2</sup>. However, in pathological conditions, numerous disorders are caused or characterized by either excessive or insufficient angiogenesis. The best known of these conditions are diseases with abnormal excessive angiogenesis such as cancer, arthritis, chronic inflammation, infectious or autoimmune diseases,

psoriasis, choroidal neovascularization, and others. Conversely, diseases characterized by insufficient angiogenesis are preeclampsia, Alzheimer disease, stroke, amyotrophic lateral sclerosis, diabetic neuropathy, peripheral artery disease, osteoporosis, and ischemic heart disease<sup>3</sup>. The development of new blood vessels is induced when the net balance of pro- and anti-angiogenic molecules is tipped in favour of angiogenesis, resulting in the ‘angiogenic switch’. During tumour growth angiogenesis is induced by a variety of stimuli. Rapid cell proliferation causes hypoxia as oxygen diffusion is limited by tissue growth. Hypoxia is an important stimulus of tumor vessel growth<sup>4</sup>, which leads to stabilization of hypoxia-inducible factor-1 $\alpha$  (HIF-1  $\alpha$ ) and HIF-1 target genes leading to angiogenesis.

## Corresponding Author

**Dr Feroz Alam**

Assistant Professor

Address:

Department of Pathology,  
Jawaharlal Nehru Medical College,  
Aligarh Muslim University  
(AMU), Aligarh, Uttar Pradesh,  
India -202002  
ferozalam97@gmail.com

## How to cite:

Alam F, Akhtar K, Maheshwari V, Hasan M, Tumor Anti-angiogenesis: Nano-Arsenal will kill the Crab. *Future Health*. 2023;01; *Future Health* 2023; 1(1):84-90.

**Submitted:** 01 June 2023

**Revised:** 28 June 2023

**Accepted:** 28 June 2023

Some key angiogenic activators include VEGF (vascular endothelial growth factor hereafter VEGF), MMPs (matrix metalloproteinases), PlGF (placenta growth factor), FGF (fibroblast growth factor) and HGF (hepatocyte growth factor)<sup>5</sup>. Endogenous inhibitors of angiogenesis include thrombospondins (THSBs), endostatin, angiostatin and cytokines such as interleukin-12<sup>6</sup>. New blood vessel formation begins with the removal of pericytes from pre-existing blood vessels, initiating the degradation of the endothelial cell basement membrane and extracellular matrix (ECM), a process regulated by the matrix metalloproteinases (MMPs). After this degradation, endothelial cells proliferate and migrate until they form unstable microvessels. Mesenchymal cells differentiate into pericytes, which allow the stability of newly formed vessels and establishing blood flow<sup>7</sup>. This process of angiogenesis is a very crucial step for tumor formation<sup>8</sup>. However, the tumor vasculature and its Anatomy are distinct compared to normal vessels - (i) tumor vessels lack the hierarchy of arterioles, capillaries and venules; (ii) tumor vasculature is disorganized and tortuous; (iii) vessels in tumors are leakier than their counterpart in normal tissues since TAECs (tumor associated ECs) are not in close contact with pericytes and are loosely connected to basement membrane; (iv) in some circumstances tumor cells may line blood vessels via vasculogenic mimicry; and (v) compared to ECs isolated from non-tumor tissues, several other molecules have been found to be enriched in TAECs indicative of their distinct molecular properties<sup>9</sup>.

**VEGF-** Tumor cells stimulate the formation of new blood vessels by means of enhanced production of the major angiogenic growth factor VEGF<sup>10</sup>, blockade of VEGF or its receptors reduces tumor growth in multiple models<sup>11</sup>. The intricacies of angiogenesis and tumor growth seem to be coordinated by cross-talk between VEGF, its receptors, and integrins<sup>12</sup>. VEGF-A is the major angiogenic growth factor and directly stimulates endothelial cells, mainly via its receptor VEGFR-2 (flk-1) or VEGFR-1 (flt-1)<sup>13</sup>. This interaction could be reverted by the monoclonal anti-VEGF antibody bevacizumab, suggesting a potential therapeutic role<sup>14</sup>.

**Integrins-** Integrins are cell surface receptors for extracellular matrix (ECM) proteins that also play a role in cell-cell attachment. Integrins are important regulators for many different cell processes including both vasculogenesis and angiogenesis<sup>15</sup>. Blood vessel formation is critically dependent on ECM, whereby integrins are the major adhesion receptors to link ECM proteins with the cytoskeleton<sup>16</sup>. Of the integrins,  $\alpha\beta3$  is one of the most extensively studied, and has an important role in angiogenesis. It binds and activates MMP-2 at the tips of growing blood vessels to help break down the ECM<sup>17</sup>. The activation of integrins can be triggered by cytokines of malignant tumor, and

blocking  $\alpha\beta3$  integrin inhibits tumor angiogenesis as well as blood vessel formation in in-vivo models<sup>18</sup>. Hence,  $\alpha\beta3$  is a potential target for tumor anti-angiogenic therapy.

**Anti-angiogenic therapy-** As already mentioned, tumor blood vessels are distinct from normal resting blood vessels, and this distinctness makes them a potential candidate for targeted cancer therapy. In order to block tumor growth and development of metastasis, a number of inhibitors targeting the tumor vasculature have been identified in different in-vitro and in-vivo studies<sup>19</sup>. Anti-angiogenic therapeutic drugs may act by, inhibiting synthesis of angiogenic proteins by cancer cells, neutralizing the angiogenic proteins, blocking the receptors for angiogenic proteins on endothelium, or directly inducing endothelial cell apoptosis. These inhibitors include therapeutic antibodies and small molecules both capable of targeting angiogenic growth factors, such as VEGF and FGF, or angiogenic growth factor receptors, such as VEGFR and PDGFR. The anti-angiogenic efficacy of chemotherapy is better observed when comparatively low doses of a chemotherapeutic agent are administered on a frequent or continuous schedule. This approach, called metronomic chemotherapy refers to the frequent administration of chemotherapeutic agents at doses significantly below the MTD (maximum tolerated dose) with no prolonged drug-free breaks<sup>20,21</sup>. Studies have also shown that the endothelial cells of newly forming blood- capillaries are highly and selectively sensitive to very low doses of various chemotherapeutic drugs<sup>22-25</sup>. Despite several advances there still remain several hindrances to treatment of cancer by anti-angiogenic therapy, including low selective toxicity of anti-cancer drugs, high tumor interstitial fluid pressure (IFP) leading to impaired transport of drug, vasculogenic mimicry in solid tumors, toxicities induced by anti-angiogenic therapy, and drug resistance. Novel strategies to overcome these problems are urgently necessary.

**Nanotechnology and anti-angiogenesis** - The advantage of nanoscale drug delivery systems is their ability to alter the pharmacokinetics and the bio-distribution of the associated therapeutic agents<sup>26</sup>. Nanotechnology having ability to engineer materials at the scale of nanometer (nm)<sup>27</sup>, nanoparticles (NPs) with at least one dimension in the range of 1 to 100 nm are widely applied in the field of medicine specially cancer. For their size NPs have unique operating ability in the complex bio-environment to interact at the level of biomolecules<sup>28</sup>. The unique size, optical, electrical, magnetic, chemical, and ligands carrying properties of nanoparticles can be targeted to the cancer cells with specificity and monitored efficiently with extreme precision in real-time<sup>29</sup>.

Nanoparticle circulation is selectively facilitated by

biological membranes<sup>30</sup>, surface modification of NPs may prevent opsonization by the reticulo-endothelial system (RES), and facilitates their retention in blood circulation; the enhanced permeability and retention (EPR) effect<sup>31</sup>. It gives advantage for using nanoparticulate contrast agents in tumor diagnostic MRI, optical imaging, photo-acoustic imaging, as well as NP delivered therapy<sup>32,33</sup>. For example, polymer-conjugated angiogenesis inhibitor TNP-470 (caplostatin) was found to accumulate selectively in the tumor vessels by the EPR effect and inhibit hyperpermeability of tumor blood vessels<sup>34</sup>. Certain nanoparticle-conjugated chemotherapeutic agents such as doxorubicin and angiogenic small molecule inhibitors have been shown to preferentially home into tumors by the EPR effect, resulting in selective vascular shutdown and inhibition of tumor growth<sup>35,36</sup>. Neutrally charged NPs of average diameter of 10–100 nm and molecular weight around 30 kDa, accumulate inside a tumor due to the EPR effect and referred to 'Passive targeting', however passive targeting may be limited in utility for its low tumor specificity, thereby, lower than required concentration at the tumor target<sup>37</sup>. Desired specificity and concentration hence, may require 'Active targeting' of the nano- vehicle with moieties such as small ligands, antibodies, and biomarkers capable of specific binding to tumor expressed molecular receptors, facilitating efficient tumor uptake, internalization and receptor-mediated endocytosis resulting in elevated concentrations in tumor cells (Figure-1)<sup>37</sup>. Monoclonal antibodies (mAbs) in their

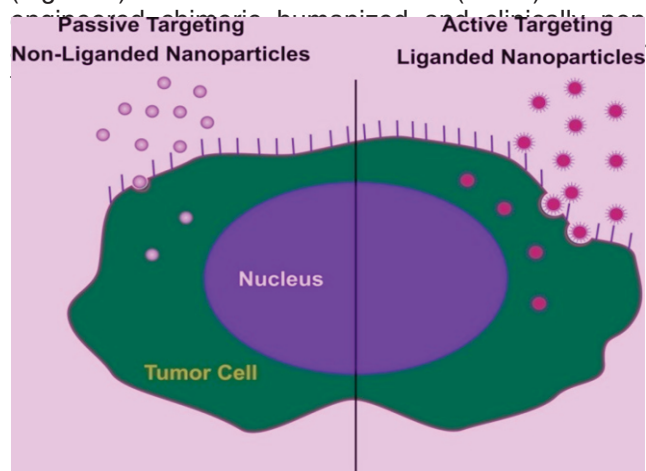


Figure 1- Showing 'Passive' and 'Active' targeting of nanoparticles

**Nanoparticle based anti-angiogenesis** - Several nanoparticles have been mediating anti-angiogenic therapy and imaging of the tumor vasculature. These include certain synthetic and few natural nanoparticles, such as polymeric conjugates and

polymeric nanoparticles like liposomes and micelles, synthetic organic nanoparticles such as dendrimers, carbon-based nanostructures such as carbon nanotubes and polyhydroxylated fullerenes, inorganic nanoparticles of gold, silver and iron-oxide, quantum dots, viral capsids and ferritin among others.

**Polymeric nanoparticles** - Polymeric nanoparticles represent one of the most effective nanocarriers for prolonged drug delivery, in the 1980s polyalkylcyanoacrylate-based nanoparticles releasing doxorubicin led to the development of polymer-based materials for drug delivery<sup>38</sup>. Langer and Folkman demonstrated the first controlled release of macromolecules using polymers, which allowed the development of anti-angiogenic drug delivery systems for cancer therapy and opened new areas for the delivery of macromolecules<sup>39</sup>. In 1994, nanoparticles composed of poly(lactic acid)/poly (lactic-co-glycolic acid) (PLA/PLGA) and PEG block copolymer were identified as "long-circulating nanoparticles" due to their stealth properties leading to an increased interest in polymeric nanoparticles and their therapeutic applications [40]. Several polymeric systems have been shown to inhibit angiogenesis and still others are under investigation.

**HPMA copolymers-** HPMA copolymers have been extensively studied for their anti-angiogenic potential. The HPMA copolymer was conjugated for the first time to the well known angiogenesis inhibitor TNP-470 (Caplostatin)<sup>41</sup>. The drug-polymer nanoconjugate selectively accumulated in the tumor microvasculature, leading to decreased tumor growth in human melanoma and lung carcinoma mice models. The drug-polymer nanoconjugate also prevented it from crossing the blood-brain barrier, and preventing neurotoxicity which is a major obstacle in the clinical use of the potent anti-angiogenic drug TNP-470. The action of Caplostatin were attributed to inhibition of various angiogenesis signaling pathways such as VEGF receptor-2 (VEGFR-2), mitogen-activated protein kinase (MAPK) and RhoA [42]. HPMA copolymers have also been used for bone targeted anti-angiogenic therapy<sup>43,44</sup>. Specific peptide sequences have been conjugated to HPMA copolymers for active targeting of the  $\alpha\beta 3$  integrin in tumor-associated vasculature. Radionuclide labeled, cyclized RGD peptide-tagged HPMA copolymer based nanoconjugates have been designed that provide the potential for targeted delivery of radionuclides and drugs to solid tumors for diagnostic and therapeutic applications<sup>45,46</sup>.

**Poly(lactic co-glycolic acid) (PLGA) copolymers** - Poly(lactic-co-glycolic acid) is a copolymer synthesized from two different monomers glycolic acid and lactic acid<sup>47</sup>. Sengupta et al. developed a PLGA based nanosystem called as 'nanocell' which was encapsulated within a polyethylene glycol (PEG)-linked lipids envelop<sup>48</sup>. PEGylation of the nanosystem renders it

nontoxic and non-immunogenic, and is an FDA approved method<sup>49</sup>. To this nanosystem, the chemotherapeutic drug doxorubicin was covalently attached to the inner PLGA core, and the anti-angiogenic agent combretastatin was trapped in the outside lipid envelope. The nanocell was destined to disrupt inside the tumor micro-environment and releasing combretastatin, which led to vascular collapse of the tumor. Local hypoxia later, led to the release of doxorubicin, which resulted in significant regression of various tumors including melanoma [48]. PLGA nanoparticles have also been used for delivering natural products thought to have anti-cancer effects. Curcumin loaded PLGA nanoparticles were reported to successfully suppress tumor necrosis factor (TNF)-regulated expression of VEGF, culminating in reduced tumor metastasis [50].

In addition, certain other polymers have been used to construct nanoparticles used to inhibit angiogenesis. Chitosan nanoparticles have shown significant inhibition of tumor growth and induction of tumor necrosis in a mouse hepatocellular carcinoma xenograft model<sup>51</sup>. Dendrimers (branched polymers) have been used to target VEGF receptors on tumor neovasculature<sup>52</sup>.

**Lipid nanoparticles** - Liposomes are self assembling, spherical, closed colloidal structures composed of lipid bilayers that surround a central aqueous space. Liposomes are able to encapsulate lipophilic or hydrophilic drugs within their lipidic layers or in their aqueous core respectively and deliver those to target site for in vivo application. Moreover, liposome delivery system can increase the solubility of hydrophobic drugs and stabilize a variety of therapeutic agents such as peptides, proteins and nucleotides in blood stream<sup>53,54</sup>. The  $\alpha\beta_3$  integrin over-expressed by endothelial cells in the tumor vasculature was targeted for drug delivery using its ligands RGD and the cyclic RGD (cRGD) [55]. RGD-targeted paclitaxel or doxorubicin-loaded PEGylated liposomes showed superior therapeutic activity over free drug or untargeted liposomes<sup>56,57</sup>. The antitumor activity of RGD-targeted paclitaxel-loaded liposomes is due to tumor microvessel destruction<sup>58</sup>. Coupling doxorubicin loaded liposomes with a peptide targeted to bombesin receptors (overexpressed in cancers) improves the therapeutic efficacy of the complex<sup>59</sup>. A peptide antagonist ATN-161 showed antineoplastic and antimetastatic properties against  $\alpha_5\beta_1$  integrin<sup>60</sup>. ATN-161 conjugated to doxorubicin-loaded PEGylated liposomes increased their therapeutic activity in a melanoma model<sup>61</sup>. Doxorubicin-loaded PEGylated liposomes functionalized with a peptide were used to target a CD13 isoform overexpressed in the tumor neovasculature<sup>62-64</sup>. Cationic liposomes can selectively bind and internalize to tumor endothelial cells due to the enrichment of the cell membranes with

negatively charged lipids and heparan sulfate proteoglycan<sup>65,66</sup>. Superior accumulation of oxaliplatin in lung tumors was obtained after intravenous injection of PEG-coated cationic drug-loaded liposomes over neutral liposomes<sup>67</sup>. Cationic liposomes have also been used for delivery of siRNA against the neoangiogenesis regulator, Argonaute 2 (Ago2) which resulted in Ago silencing in tumors together with apoptosis of tumor blood vessels and decreased tumor growth [68,69]. In another study, paclitaxel-loaded cationic liposomes (EndoTAG-1) induced endothelial cell apoptosis in vivo, retarded melanoma and pancreatic carcinoma tumor growth, and decreased the number of melanoma lung metastases in vivo<sup>70-72</sup>. Targeting of tumor vasculature by an aptamer directed against the tumor vasculature marker E-selectin has also been reported<sup>73,74</sup>. Coupling of a peptide ligand to doxorubicin-loaded liposomes increased doxorubicin accumulation in neuroblastoma leading to destruction of perivascular and endothelial cells and significant increase in survival of neuroblastoma bearing mice over either endothelial cell targeted or pericyte-targeted liposomes alone<sup>75</sup>.

Other lipid based nanoparticles used for cancer therapy are the micelles having a core-shell structure. The core of the micelles, which is either the hydrophobic part or the ionic part, can contain small (or bigger) molecules such as therapeutic drugs. The shell provides interactions with the solvent and make the nanoparticles stable in the liquid<sup>76</sup>. Given their lipophilic nature, most anticancer drugs are inherently water insoluble. By encapsulation of the drug within the hydrophobic core of the micelle, the apparent solubility of the drug can be significantly increased. Hence, micelles allow for the in vivo use of drugs otherwise deemed too hydrophobic or toxic, without having to manipulate the chemical structure of the agent. Additionally, encapsulating the drug within the polymer core affords drug stability by hindering enzymatic degradation and inactivation. Lodamin a novel nano-formulation was developed with the use of poly(ethylene-glycol)-poly(lactic)acid (PEG-PLA) molecules and a very potent anti-angiogenic drug TNP-470 forming a nano-micelle complex which overcame the problems of neurotoxicity, poor bio-availability and short half-life that were associated with the use of this powerful anti cancer drug<sup>77</sup>. In another study cRGD was conjugated to the outer shell of doxorubicin-loaded polymeric micelles and these modified micelles significantly enhanced their internalization (up to 30-fold) by receptor-mediated endocytosis in tumor endothelial cells overexpressing the  $\alpha_v\beta_3$  integrin<sup>78</sup>. A micelle based delivery of Hypoxia-inducible factor-1 $\alpha$  siRNA (HIF-1 $\alpha$  siRNA) was used to treat hypoxic tumors<sup>79</sup>.

**Carbon nanoparticles** - Carbon based nanoparticles like nanotubes, nanofibers and fullerenes have gained

considerable attention in the field of medicine. Carbon nanotubes (CNTs) are the most studied one, having a basic structure of rolled up seamless cylinders of graphene sheets consisting of a honeycomb of benzene rings in the same plane. Depending on the number of graphene layers from which a single nanotube is composed, CNTs are classified as single-walled carbon nanotubes (SWNTs) and multi-walled carbon nanotubes (MWNTs) [80]. CNT-based drug delivery has shown promise for the intracellular delivery of small drug molecules, DNA plasmids, short-interfering RNA (siRNA), and proteins, for cancer therapies *in vitro* and *in vivo* [81]. Carbon nanomaterials are being increasingly studied for their anti-angiogenic potential. Murugesan et al. reported the anti-angiogenic potential of pristine (unmodified) MWNTs, C60 fullerenes and graphite in chick chorioallantoic membrane (CAM) by inhibiting VEGF- and bFGF-induced angiogenesis<sup>82</sup>. However, these carbon materials did not show any significant effect on basal angiogenesis in the absence of added growth factors, indicating their differential anti-angiogenic potential only in tumor environment where angiogenic factors are known to be upregulated. In another study, Chaudhuri et al evaluated the effect of doxorubicin conjugated single walled carbon nanotubes (CNT-Dox) and doxorubicin-conjugated spherical polyhydroxylated fullerenes or fullereneols (Ful-Dox) on angiogenesis<sup>83</sup>. They reported that CNTs exert a pro-angiogenic effect *in vitro* and *in vivo*. In contrast, the fullereneols or doxorubicin-conjugated fullereneols exerted a dramatically opposite antiangiogenic activity in zebrafish and murine tumor angiogenesis models. Other studies have also used carbon nanomaterials for evaluating their anti-angiogenic potential<sup>84,85</sup>.

**Inorganic nanoparticles-** Several inorganic nanoparticles, such as those of gold, silver and iron-oxide, possess unique properties, which are increasingly being utilized for biomedical applications, especially cancer diagnosis and treatment. Studies have also demonstrated the anti-angiogenic potential of these inorganic nanoparticles. Bartczak et al. demonstrated deliberate activation or inhibition of *in vitro* angiogenesis using functional peptide coated gold nanoparticles. The functional particles were shown to influence the extent and morphology of vascular structures, without causing toxicity. Mechanistic studies showed that the nanoparticles have the ability to alter the balance between naturally secreted pro- and anti-angiogenic factors, under various biological conditions [86]. Studies have also reported similar anti-angiogenic properties of silver nanoparticles<sup>87-89</sup>.

## Conclusions

Tumor angiogenesis in recent times has been used as

a potential target for therapy and the effects of anti-angiogenic therapy has improved the therapeutic index of cancer chemotherapy. The development of targeted cancer therapeutics with improved ability to discriminate between tumor cells and normal cells is one of the major goals of current anti-cancer research. Nanotechnology has presented many innovative methodologies being utilized for targeted drug delivery with minimal toxicities. Hopefully, in future a combinatorial approach of nanotechnology and anti-angiogenesis will lead to the development of such drugs which will destroy a large tumor in very short duration with only minimal discomfort and side-effects to the patient.

## References

1. Karamysheva AF. Mechanisms of Angiogenesis. *Biochemistry (Mosc)*. 2008; 73: 751-762.
2. Risau W, Flamme I. Vasculogenesis. *Annu Rev Cell Dev Biol* 1995;11:73–91.
3. Carmeliet P. Angiogenesis in health and disease. *Nat Rev* 2003;9:653-660.
4. Dang DT, Chun SY, Burkitt K, Abe M, Chen S, Havre P et al. Hypoxia-inducible factor-1 target genes as indicators of tumor vessel response to vascular endothelial growth factor inhibition. *Cancer Res* 2008; 68: 1872–1880.
5. Khoury CC, Ziyadeh FN. Angiogenic factors. *Contrib Nephrol* 2011; 170: 83–92.
6. Tarabozetti G, Rusnati M, Ragona L, Colombo G. Targeting tumor angiogenesis with TSP-1-based compounds: rational design of antiangiogenic mimetics of endogenous inhibitors. *Oncotarget* 2010;1: 662–673.
7. Nussenbaum F, Herman I M. Tumor angiogenesis: insights and innovations. *J Oncol*. 2010; 2010: 132641. <https://doi.org/10.1155/2010/132641>
8. Folkman J. Tumor angiogenesis: therapeutic implications. *N Engl J Med* 1971; 285 :1182-1186.
9. S. Hiratsuka. Vasculogenesis, angiogenesis and special features of tumor blood vessels. *Front Biosci* 2011;16:1413–1427.
10. Folkman J. Role of angiogenesis in tumor growth and metastasis. *Semin Oncol* 2002; 29: 15–18.
11. Ferrara N. Vascular endothelial growth factor: basic science and clinical progress. *Endocr Rev* 2004; 25: 581–611.
12. Varner JA & Cheresch DA. In *Important Advances in Oncology*: 1996, eds. Devita, V. T., Hellman, S. & Rosenberg, S. A. (Lippincott Williams & Wilkins, Baltimore), pp. 69–87.
13. Kendall RL, Wang G, DiSalvo J, Thomas KA. Specificity of vascular endothelial cell growth factor receptor ligand binding domains. *Biochem Biophys Res Commun* 1994; 201: 326–330.
14. Prager GW, Lackner EM, Krauth MT, Unseld M, Poettler M, Laffer S et al. Targeting of VEGF-dependent

- transendothelial migration of cancer cells by bevacizumab. *Mol Oncol* 2010; 4:150–160.
15. Avraamides CJ, Garmy-Susini B, Varner JA. Integrins in angiogenesis and lymphangiogenesis. *Nat Rev Cancer* 2008; 8:604-617.
  16. Friedlander M, Brooks PC, Shaffer RW, Kincaid CM, Varner JA, Cheresh DA et al. Definition of two angiogenic pathways by distinct alpha v integrins. *Science* 1995; 270: 1500–1502.
  17. Vartak DG, Lee BS, Gemeinhart RA. In vitro evaluation of functional interaction of integrin alphavbeta3 and matrix metalloprotease-2. *Mol Pharm.* 2009 6:1856-1867.
  18. Drake CJ, Cheresh DA, Little CD. An antagonist of integrin alpha v beta 3 prevents maturation of blood vessels during embryonic neovascularization. *J Cell Sci* 1995; 108: 2655–2661.
  19. Ferrara N, Kerbel RS. Angiogenesis as a therapeutic target. *Nature* 2005; 438: 967-974.
  20. Browder T, Butterfield CE, Kraling BM, Shi B, Marshall B, O'Reilly MS, et al. Antiangiogenic scheduling of chemotherapy improves efficacy against experimental drug resistant cancer. *Cancer Res* 2000; 60: 1878-1886.
  21. Hanahan D, Bergers G, Bergsland E. Less is more, regularly: metronomic dosing of cytotoxic drugs can target tumor angiogenesis in mice. *J Clin Invest* 2000; 105: 1045-1047.
  22. Belotti D, Vergani V, Drudis T, Borsotti P, Pitelli MR, Viale G, et al. The microtubule-affecting drug paclitaxel has antiangiogenic activity. *Clin Cancer Res* 1996; 2: 1843-1849.
  23. Bocci G, Nicolaou KC, Kerbel RS. Protracted low-dose effects on human endothelial cell proliferation and survival in vitro reveal a selective antiangiogenic window for various chemotherapeutic drugs. *Cancer Res* 2002; 62: 6938-6943.
  24. Vacca A, Iurlaro M, Ribatti D, Minischetti M, Nico B, Ria R, et al. Antiangiogenesis is produced by nontoxic doses of vinblastine. *Blood* 1999; 94: 4143-4155.
  25. Wang J, Lou P, Lesniewski R, Henkin J. Paclitaxel at ultra low concentrations inhibits angiogenesis without affecting cellular microtubule assembly. *Anticancer Drugs* 2003; 14: 13-19.
  26. Northfelt DW, Martin FJ, Working P, Volberding PA, Russell J, Newman M et al. Doxorubicin encapsulated in liposomes containing surface-bound polyethylene glycol: pharmacokinetics, tumor localization, and safety in patients with AIDS-related Kaposi's sarcoma. *J Clin Pharmacol* 1996; 36: 55-63.
  27. Ramsden JJ. What is Nanotechnology?. *Nanotechnology perceptions Journal (Collegium Basilea)* 2005; 1:3-17.
  28. McNeil SE, Nanotechnology for the biologist. *J Leukoc Biol* 2005; 78:585-594.
  29. Yang F, Jin C, Subedi S, Lee CL, Wang Q, Jiang Y, et al. Emerging inorganic nanomaterials for pancreatic cancer diagnosis and treatment. *Cancer Treat Rev* 2012; 38:566-579.
  30. Longmire M, Choyke PL, Kobayashi H. Clearance properties of nano-sized particles and molecules as imaging agents: Considerations and caveats. *Nanomedicine (Lond)* 2008; 3:703-717.
  31. Chatterjee DK, Fong LS, Zhang Y. Nanoparticles in photodynamic therapy: An emerging paradigm. *Adv Drug Deliv Rev* 2008;60:1627-1637.
  32. Jain RK. Lessons from multidisciplinary translational trials on anti-angiogenic therapy of cancer. *Nat Rev Cancer* 2008; 8:309-316.
  33. Iyer AK, Khaled G, Fang J, Maeda H. Exploiting the enhanced permeability and retention effect for tumor targeting, *Drug Discov Today* 2006;11:812–818.
  34. Satchi-Fainaro R, Puder M, Davies JW, Tran HT, Sampson DA, Greene AK, et al. Targeting angiogenesis with a conjugate of HPMA copolymer and TNP-470. *Nat Med* 2004; 10:255-261.
  35. Guo Q, Wang H, Zhao Y, Wang H, Zeng F, Hua H et al. Cell-penetrating albumin conjugates for enhanced doxorubicin delivery. *Polym Chem* 2013;4: 4584-4587.
  36. Chaudhuri P, Harfouche R, Soni S, Hentschel DM, Sengupta S. Shape effect of carbon nanovectors on angiogenesis. *ACS Nano* 2010; 4:574-582.
  37. Fernandez-Fernandez A, Manchanda R, McGoron AJ. Theranostic applications of nanomaterials in cancer: Drug delivery, image-guided therapy, and multifunctional platforms. *Appl Biochem Biotechnol* 2011; 165:1628-1651.
  38. Couvreur P, Kante B, Roland M, Speiser P. Adsorption of antineoplastic drugs to polyalkylcyanoacrylate nanoparticles and their release in calf serum. *J Pharm Sci* 1979; 68:1521– 1524.
  39. Langer R, Folkman J. Polymers for the sustained release of proteins and other macromolecules. *Nature* 1976; 263:797–800.
  40. Gref R, Minamitake Y, Peracchia MT, Trubetskoy V, Torchilin V, Langer R. Biodegradable long-circulating polymeric nanospheres. *Science* 1994; 263:1600–1603.
  41. Satchi-Fainaro R, Puder M, Davies JW, Tran HT, Sampson DA, Greene AK, et al. Targeting angiogenesis with a conjugate of HPMA copolymer and TNP-470. *Nat Med* 2004;10:255-261.
  42. Satchi-Fainaro R, Mamluk R, Wang L, Short SM, Nagy JA, Feng D, et al. Inhibition of vessel permeability by TNP-470 and its polymer conjugate, caplostatin. *Cancer Cell* 2005;7:251-261.
  43. Segal E, Pan H, Ofek P, Udagawa T, Kopeckova P, Kopecek J, et al. Targeting angiogenesis-dependent calcified neoplasms using combined polymer therapeutics. *PLoS ONE* 2009, 4:e5233.
  44. Miller K, Erez R, Segal E, Shabat D, Satchi-Fainaro R. Targeting bone metastases with a bispecific anticancer and antiangiogenic polymeralendronate- taxane conjugate. *Angew Chem Int Ed Engl* 2009; 48:2949-2954.
  45. Mitra A, Mulholland J, Nan A, McNeill E, Ghandehari H,

- Line BR. Targeting tumor angiogenic vasculature using polymer-RGD conjugates. *J Control Release* 2005; 102:191-201.
46. Mitra A, Coleman T, Borgman M, Nan A, Ghandehari H, Line BR. Polymeric conjugates of mono- and bi-cyclic alphaVbeta3 binding peptides for tumor targeting. *J Control Release* 2006; 114:175-183.
  47. Astete CE, Sabliov CM. Synthesis and characterization of PLGA nanoparticles. *J. Biomater. Sci. Polym. Ed.* 2006;17:247–289.
  48. Sengupta S, Eavarone D, Capila I, Zhao G, Watson N, Kiziltepe T, Sasisekharan R. Temporal targeting of tumour cells and neovasculature with a nanoscale delivery system. *Nature* 2005; 436:568-572.
  49. Veronese FM, Pasut G. PEGylation, successful approach to drug delivery. *Drug Discov Today* 2005; 10:1451-1458.
  50. Anand P, Nair HB, Sung B, Kunnumakkara AB, Yadav VR, Tekmal RR, et al. Design of curcumin-loaded PLGA nanoparticles formulation with enhanced cellular uptake, and increased bioactivity in vitro and superior bioavailability in vivo. *Biochem Pharmacol* 2010; 79:330-338.
  51. Xu Y, Wen Z, Xu Z. Chitosan nanoparticles inhibit the growth of human hepatocellular carcinoma xenografts through an antiangiogenic mechanism. *Anticancer Res* 2009; 29:5103-5109.
  52. Backer MV, Gaynutdinov TI, Patel V, Bandyopadhyaya AK, Thirumamagal BT, Tjarks W et al. Vascular endothelial growth factor selectively targets boronated dendrimers to tumor vasculature. *Mol Cancer Ther* 2005; 4:1423-1429.
  53. Irache JM, Esparza I, Gamazo C, Agüeros M, Espuelas S. Nanomedicine: novel approaches in human and veterinary therapeutics. *Vet Parasitol* 2011; 180: 47-71.
  54. Allen TM, Moase EH. Therapeutic opportunities for targeted liposomal drug delivery. *Adv Drug Deliv Rev* 1996; 21:117-133.
  55. Danhier F, Breton AL, and Preat V. RGD-based strategies to target alpha(v) beta(3) integrin in cancer therapy and diagnosis. *Mol. Pharmaceutics* 2012; 9:2961–2973.
  56. Zhao H, Wang JC, Sun QS, Luo CL and Zhang Q. RGD-based strategies for improving antitumor activity of paclitaxel-loaded liposomes in nude mice xenografted with human ovarian cancer. *J Drug Target* 2009; 17: 10–8. doi: 10.1080/10611860802368966.
  57. Xiong XB, Huang Y, Lu WL, Zhang X, Zhang H, Nagai T. Intracellular delivery of doxorubicin with RGD-modified sterically stabilized liposomes for an improved antitumor efficacy: invitro and invivo. *J Pharm Sci* 2005; 94: 1782–1793.
  58. Meng S, Su B, Li W, Ding Y, Tang L, Zhou W et al. Integrin-targeted paclitaxel nanoliposomes for tumor therapy. *Med Oncol* 2011; 28: 1180–1187.
  59. Accardo A, Salsano G, Morisco A, Aurilio M, Parisi A, Maione F et al. Peptide-modified liposomes for selective targeting of bombesin receptors overexpressed by cancer cells: a potential theranostic agent. *Int J Nanomedicine* 2012; 7: 2007–2017.
  60. Doñate F, Parry GC, Shaked Y, Hensley H, Guan X, Beck I et al. Pharmacology of the novel antiangiogenic peptide ATN-161 (Ac-PHSCN-NH<sub>2</sub>): observation of a U-shaped dose-response curve in several preclinical models of angiogenesis and tumor growth. *Clin Cancer Res* 2008; 14: 2137–2144.
  61. Dai W, Yang T, Wang Y, Wang X, Wang J, Zhang X et al. Peptide PHSCNK as an integrin alpha(5)beta(1) antagonist targets stealth liposomes to integrin overexpressing melanoma. *Nanomedicine* 2012; 8: 1152–1161.
  62. Pastorino F, Di Paolo D, Piccardi F, Nico B, Ribatti D, Daga A et al. Enhanced antitumor efficacy of clinical-grade vasculature-targeted liposomal doxorubicin. *Clin Cancer Res* 2008; 14: 7320–7329.
  63. Takara K, Hatakeyama H, Kibria G, Ohga N, Hida K, Harashima H. Size-controlled, dual-ligand modified liposomes that target the tumor vasculature show promise for use in drug resistant cancer therapy. *J Control Release* 2012; 162: 225–232.
  64. Colombo G, Curnis F, De Mori GMS, Gasparri A, Longoni C, Sacchi A et al. Structure activity relationships of linear and cyclic peptides containing the NGR tumor-homing motif. *J Biol Chem* 2002; 277: 47891–47899.
  65. Thurston G, McLean JW, Rizen M, Baluk P, Haskell A, Murphy TJ et al. Cationic liposomes target angiogenic endothelial cells in tumors and chronic inflammation in mice. *J Clin Invest* 1998; 101:1401–1413.
  66. Ran S and Thorpe PE. Phosphatidylserine is a marker of tumor vasculature and a potential target for cancer imaging and therapy. *Int J Radiat Oncol Biol Phys* 2002; 54:1479–1484.
  67. Abu Lila AS, Kizuki S, Doi Y, Suzuki T, Ishida T, and H. Kiwada. Oxaliplatin encapsulated in PEG-coated cationic liposomes induces significant tumor growth suppression via a dual-targeting approach in a murine solid tumor model. *J Control Release* 2009; 137: 8–14.
  68. Tagami T, Suzuki T, Matsunaga M, Nakamura K, Moriyoshi N, Ishida T et al. Anti-angiogenic therapy via cationic liposome-mediated systemic siRNA delivery. *Int J Pharm* 2012; 422: 280–289.
  69. Asai T, Suzuki Y, Matsushita S, Yonezawa S, Yokota J, Katanasaka Y et al. Disappearance of the angiogenic potential of endothelial cells caused by Argonaute2 knockdown. *Biochem Biophys Res Commun* 2008; 368: 243–248.
  70. Eichhorn ME, Becker S, Strieth S, Werner A, Sauer B, Teifel M et al. Paclitaxel encapsulated in cationic lipid complexes (MBT-0206) impairs functional tumor vascular properties as detected by dynamic contrast enhanced magnetic resonance imaging. *Cancer Biol Ther* 2006; 5: 89–96.
  71. Schmitt-Sody M, Strieth S, Krasnici S, Sauer B, Schulze B, Teifel M. Neovascular targeting therapy: paclitaxel encapsulated in cationic liposomes improves antitumoral efficacy. *Clin Cancer Res* 2003; 9: 2335–2341.

72. Bode C, Trojan L, Weiss C, Kraenzlin B, Michaelis U, Teifel M. Paclitaxel encapsulated in cationic liposomes: a new option for neovascular targeting for the treatment of prostate cancer. *Oncol Rep* 2009; 22: 321–326.
73. Mann AP, Bhavane RC, Somasunderam A, Liz Montalvo-Ortiz B, Ghaghada KB, Volk D et al. Thioaptamer conjugated liposomes for tumor vasculature targeting. *Oncotarget* 2011; 2: 298–304.
74. Marchiò S, Lahdenranta J, Schlingemann RO, Valdembri D, Wesseling P, Arap MA. Aminopeptidase A is a functional target in angiogenic blood vessels. *Cancer Cell* 2004; 5:151–162.
75. Loi M, Marchio S, Becherini P, Di Paolo D, Soster M, Curnis F et al. Combined targeting of perivascular and endothelial tumor cells enhances anti-tumor efficacy of liposomal chemotherapy in neuroblastoma. *J Control Release* 2010; 145:66–73.
76. Torchilin VP. Micellar nanocarriers: pharmaceutical perspectives. *Pharm Res* 2007; 24:1–16.
77. Benny O, Fainaru O, Adini A, Cassiola F, Bazinet L, Adini I, et al. An orally delivered small-molecule formulation with antiangiogenic and anticancer activity. *Nat Biotechnol* 2008, 26:799-807.
78. Nasongkla N, Shuai X, Ai H, Weinberg BD, Pink J, Boothman DA, et al. cRGD-functionalized polymer micelles for targeted doxorubicin delivery. *Angew Chem Int Ed Engl* 2004, 43:6323-6327.
79. Liu XQ, Xiong MH, Shu XT, Tang RZ and Wang J. Therapeutic Delivery of siRNA Silencing HIF-1 Alpha with Micellar Nanoparticles Inhibits Hypoxic Tumor Growth. *Mol Pharmaceutics* 2012; 9: 2863–2874.
80. Usui Y, Haniu H, Tsuruoka S, Saito N. Carbon Nanotubes Innovate on Medical Technology. *Med Chem* 2012; 2: 001-006.
81. Wang X J, Liu Z. Carbon nanotubes in biology and medicine: An overview. *Chin Sci Bull* 2012; 57: 167-180.
82. Murugesan S, Mousa SA, O'connor LJ, Lincoln DW, Linhardt RJ. Carbon inhibits vascular endothelial growth factor-and fibroblast growth factor promoted angiogenesis. *FEBS Lett* 2007; 581:1157-1160.
83. Chaudhuri P, Harfouche R, Soni S, Hentschel DM, Sengupta S. Shape effect of carbon nanovectors on angiogenesis. *ACS Nano* 2010; 4:574-582.
84. Wierzbicki M, Sawosz E, Grodzik M, Prasek M, Jaworski S and Chwalibog A. Comparison of anti-angiogenic properties of pristine carbon nanoparticles. *Nanoscale Research Letters* 2013 8:195. <http://www.nanosca-lereslett.com/content/8/1/195>.
85. Chen C, Zhang H, Hou L, Shi J, Wang L, Zhang C et al. Single-Walled Carbon Nanotubes Mediated Neovascularity Targeted Antitumor Drug Delivery System. *J Pharm Pharmaceut Sci* 2013;16: 40–51.
86. Bartczak D, Muskens OL, Sanchez-Elsner T, Kanaras AG and Millar TM. Manipulation of in Vitro Angiogenesis Using Peptide-Coated Gold Nanoparticles. *ACS Nano* 2013;7:5628-5636.
87. Gurunathan S, Lee KJ, Kalishwaralal K, Sheikpranbabu S, Vaidyanathan R, Eom SH. Antiangiogenic properties of silver nanoparticles. *Biomaterials* 2009; 30:6341-6350.
88. Will SEA, Favaron PO, Pavez MA, Florentino LC, Soares D, Oliveira FC et al. Bactericidal silver nanoparticles present an antiangiogenic effect in the Chorioallantoic Membrane Model (CAM) in Science